

N-ETHOXYCARBONYL-L-PROLINAMIDE, A NEW ALKALOID FROM THE LEAVES OF *Arnica montana* L.*

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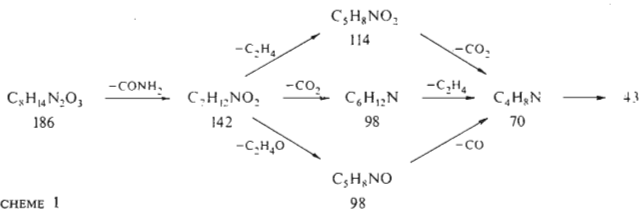
From the chloroform extract of the leaves of *Arnica montana* L. the as yet undescribed N-ethoxycarbonyl-L-prolinamide (*V*) was isolated in addition to other substances. Its structure was elucidated on the basis of mass, infrared and ¹H-NMR spectroscopy and confirmed by synthesis.

Several years ago we studied the compounds of the light petroleum extract of the leaves of *Arnica montana* L. (*Compositae*, tribe *Senecioneae*)¹. We also investigated the substances which we obtained by chloroform extraction from a material previously extracted with light petroleum. The chloroform extract was worked up similarly as the ethanolic extract obtained during the isolation of salonitenolide², and we isolated from it both substances which were also present in the light petroleum extract, such as arnicolide A (*I*), tetrahydrohelenalin (*II*) and dihydrohelenalin (*III*), and also loliolide (*IV*) described in another paper³ and, finally, an alkaloid (*V*) of the composition C₈H₁₄N₂O₃ with two active hydrogens. The infrared spectrum of substance *V* contained absorption bands at 3390, 3205, 1675 and 1628 cm⁻¹, corresponding to the amide group, and bands at 1683 and 1184, belonging to the ethoxycarbonyl group.

The structure of the substance under investigation was deduced from the following facts: Its elemental analysis agrees with the high resolution mass spectrometric determination of the molecular peak. The integral of the ¹H-NMR spectrum corresponds to 12 protons where 2 protons, unobserved in the spectrum, are exchangeable, as indicated by the number of active hydrogens. The loss of CH₂NO₂ from M⁺ and the composition of the base-peak in the mass spectrum (C₄H₈N) (Scheme 1) indicates the presence of a carboxamide group and a pyrrolidine ring in the investigated molecule. Hence, one of the alternative structure *V* and *VI* might belong to the

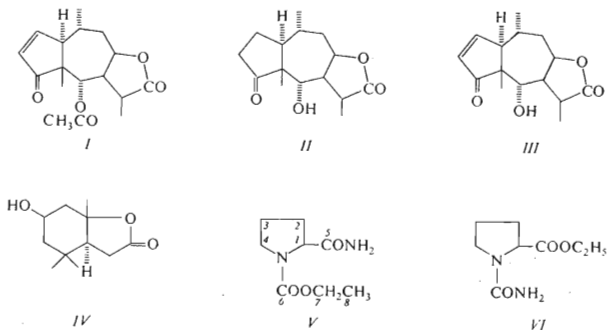
* Part XLI in the series Plant Substances; Part XL: This Journal 40, 3731 (1975).

native substance. In view of the fact that 1,2-disubstituted pyrrolidine derivatives lose on electron impact primarily substituents in the position α to the nitrogen atom^{4,5}, and in view of the fact that the fragmentation of the substance investigated agrees with the fragmentation of N-ethoxycarbonyl-L-prolinol, that begins with the ion m/e 132 (ref.⁵), we consider the structures *V* as more probable.



SCHEME 1

A definite decision as to which of the two structures is correct was brought by the synthesis of compound *V* from L-proline. Applying a known procedure⁶ we synthesized the amide of L-proline and submitted it to the reaction with ethyl chloroformate which gave N-ethoxycarbonyl-L-prolinamide (*V*). The latter was identical with the native substance in all respects. Simultaneously the synthesis of compound *V* from L-proline also proves its absolute configuration.



Up to now, N-ethoxycarbonyl-L-prolinamide (*V*) has not been found in natural material. In view of its structure this compound may be classified among the relatively scarce pyrrolidine alkaloids^{7,8}, from which it differs, however, by the substitution

with the ethoxycarbonyl group on the nitrogen atom of the pyrrolidine ring, considering that such a substitution is not common even in alkaloids of other types with an amide group.

EXPERIMENTAL

The melting points were determined on a Kofler block and they are not corrected. For column chromatography silica gel according to Pitra and Štěrba⁹ (30–60 μ , deactivated by the addition of 11% of water) was used. For thin-layer chromatography silica gel G Merck was used. The IR spectra were measured on a Unicam SP 200 and on a Zeiss UR-10 (Jena) spectrophotometer. ¹H-NMR spectra were measured on a Tesla BS 487 (80 MHz) instrument. The mass spectra were measured on a Varian MAT-311 spectrograph. Optical rotation was determined with an objective polarimeter in methanol. Circular dichroism was measured on a Roussel-Jouan Dichrograph CD 185 in methanol.

Working up of the Chloroform Extract

Dry ground leaves of *A. montana* L. (16 kg) were extracted with light petroleum¹ and then with chloroform. The chloroform extract was evaporated and the residue partitioned between light petroleum and 60% aqueous ethanol as described earlier². A residue (14.1 g) was obtained which was dissolved in benzene and then chromatographed on silica gel (300 g). The course of chromatography is summarized in Table I.

Arnicolide A (I): From fraction 1 (Table I) arnicolide A (I) was obtained by chromatography on silica gel. It was identical in all respects with a sample of arnicolide A (I) isolated from the light petroleum extract¹.

Loliolide (IV): From fraction 2 (Table I) loliolide (IV) was isolated by chromatography on silica gel, as described earlier³.

Tetrahydrohelenalin (II) and *dihydrohelenalin* (III): From fraction 2 (Table I) tetrahydrohelenalin (II) and dihydrohelenalin (III) were isolated by chromatography on silica gel; both substances were identical in all respects with those isolated from the light petroleum extract¹.

TABLE I
Chromatography of the Chloroform Extract

Fraction	Solvent	Volume, l	Residue, g	Substance
1	benzene-ether 4 : 1	1.5	1.9	I
2	benzene-ether 4 : 1	4.4	2.0	II-IV
3	benzene-ether 1 : 1	2.5	1.1	—
4	ether-methanol 9 : 1	2.0	3.9	V

N-Ethoxycarbonyl-L-prolinamide (*V*)

a) Fraction 4 (Table I; 3.8 g) was rechromatographed on silica gel (200 g). Using light petroleum, acetone and ether (4 : 3 : 3) for elution alkaloid *V* was isolated, m.p. 103–103.5°C (ether), $[\alpha]_D^{20} = -55.8^\circ$ (*c* 0.12). ORD spectrum: 350 nm, $[\Phi] = -583$; 233 nm, $[\Phi] = -3457$; 225 nm, $[\Phi] = -2684$ (through). CD spectrum: 215 nm, $\Delta\epsilon = -1.69$. IR spectrum (cm^{-1}): 3390, 3205, 1675, 1628 (CONH_2), 1685, 1184 (COOC_2H_5). Mass spectrum (*m/e*, relative intensity, composition): 186, 1.4, $\text{C}_8\text{H}_{14}\text{N}_2\text{O}_3$ (M^+); 168, 0.6; 142, 81.9, $\text{C}_7\text{H}_{12}\text{NO}_2$; 114, 11.8, $\text{C}_5\text{H}_8\text{NO}_2$; 98, 20.5, $\text{C}_6\text{H}_{12}\text{N}$, $\text{C}_5\text{H}_8\text{NO}$; 70, 100, $\text{C}_4\text{H}_8\text{N}$; 69, 15.7, $\text{C}_4\text{H}_7\text{N}$; 68, 26.8, $\text{C}_4\text{H}_6\text{N}$; 55, 6.4; 44, 22.8, CO_2 , CONH_2 , 43, 29.1; 42, 12.6; 41, 27.6. $^1\text{H-NMR}$ spectrum (chemical shifts in δ -scale, p.p.m.): 1.31, t, 3 H_8 ; $J = 7.5$ Hz; 1.85–2.45, m, 4 $\text{H}_2 + \text{H}_3$; 3.57, t, 2 H_4 , $J = 5.5$ Hz; 4.25, q, 2 H_7 , $J = 7.5$ Hz, 4.46 q, 1 H_1 , $J_1 = 5$, $J_2 = 7$. For $\text{C}_8\text{H}_{14}\text{N}_2\text{O}_3$ (186.2) calculated: 51.60% C, 7.58% H, 15.05% N; 1.08% H act.; found: 51.68% C, 7.58% H, 15.13% N, 1.12% H act.

b) Ethyl chloroformate (1.0 ml) was added in several portions to an ice-cooled solution of the amide of L-proline⁶ (1.0 g) in 5 ml 50% aqueous acetone and 1.4 ml of triethylamide, under stirring, and the mixture was stirred for an additional 3 hours at room temperature. The mixture was acidified with dilute HCl and extracted with chloroform; the combined chloroform extracts were worked up to give 430 mg of the product from which pure amide *V* (200 mg) was obtained by crystallization. M.p. 102–103°C, $[\alpha]_D^{20} = -58.1^\circ$ (*c* 0.45). The mixed melting point of the synthetic and native N-ethoxycarbonyl-L-prolinamide was undepressed. The identity is further confirmed by the identity of all physical constants measured, as well as the analytical data given under a).

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REFERENCES

1. Poplawski J., Holub M., Samek Z., Herout V.: *This Journal* 36, 2189 (1971).
2. Suchý M., Samek Z., Herout V., Šorm F.: *This Journal* 32, 2016 (1967).
3. Holub M., Samek Z., Poplawski J.: *Phytochemistry* 14, 1659 (1975).
4. Biemann K., Seibl J., Gapp F.: *J. Amer. Chem. Soc.* 83, 3795 (1961).
5. Wiegrebe W., Herrmann E. G., Schlunger U. P., Budzikiewicz H.: *Helv. Chim. Acta* 57, 301 (1974.)
6. Stewart F. H. C.: *Australian J. Chem.* 22, 2451 (1969).
7. Marion L. in the book: *The Alkaloids* (R. H. F. Manske, H. L. Homes, Eds), Vol. I, p. 91. Academic Press, New York 1950.
8. Marion L. in the book: *The Alkaloids* (R. H. F. Manske, Ed.), Vol. VI, p. 31. Academic Press, New York, London 1960.
9. Pitra J., Štěrba J.: *Chem. Listy* 56, 544 (1962).

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